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Effects of diesel exposure on lung function and inflammation biomarkers from airway and peripheral blood of healthy volunteers in a chamber study

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Abstract

Background: Exposure to diesel exhaust causes inflammatory responses. Previous controlled exposure studies at a concentration of 300 $\mu\text{g}/\text{m}^3$ of diesel exhaust particles mainly lasted for 1 h. We prolonged the exposure period and investigated how quickly diesel exhaust can induce respiratory and systemic effects.

Methods: Eighteen healthy volunteers were exposed twice to diluted diesel exhaust (PM_{10} $\sim 300 \mu\text{g}/\text{m}^3$) and twice to filtered air (PM_{10} $\sim 2 \mu\text{g}/\text{m}^3$) for 3 h, seated, in a chamber with a double-blind set-up. Immediately before and after exposure, we performed a medical examination, spirometry, rhinometry, nasal lavage and blood sampling. Nasal lavage and blood samples were collected again 20 h post-exposure. Symptom scores and peak expiratory flow (PEF) were assessed before exposure, and at 15, 75, and 135 min of exposure.

Results: Self-rated throat irritation was higher during diesel exhaust than filtered air exposure. Clinical signs of irritation in the upper airways were also significantly more common after diesel exhaust exposure (odds ratio=3.2, $p < 0.01$). PEF increased during filtered air, but decreased during diesel exhaust exposure, with a statistically significant difference at 75 min (+4 L/min vs. -10 L/min, $p = 0.005$). Monocyte and total leukocyte counts in peripheral blood were higher after exposure to diesel exhaust than filtered air 20 h post-exposure, and a trend ($p = 0.07$) towards increased serum IL-6 concentrations was also observed 20 h post-exposure.

Conclusions: Diesel exhaust induced acute adverse effects such as symptoms and signs of irritation, decreased PEF, inflammatory markers in healthy volunteers. The effects were first seen at 75 min of exposure.

Keywords: Diesel exhaust, Chamber experiment, Lung function, Biomarkers, Respiratory inflammation

Background

The adverse health effects of particulate air pollution are well documented [1-5], and urban outdoor air pollution was estimated to cause 1.34 million premature deaths worldwide in 2008, an increase of 16% from 2004 [6]. Diesel exhaust (DE) emissions constitute, according to estimates from the USA, 4-16% of the total particulate matter (PM) in non-urban and urban areas [7,8]. People working around diesel equipment or living close to major

roadways are more likely to have irritant respiratory symptoms and reduced baseline pulmonary function [9,10], and to develop respiratory disease including allergy [11], chronic obstructive pulmonary disease [12,13], and lung cancer [14-16].

Epidemiological studies have shown associations between DE and respiratory diseases, but the underlying mechanisms are still not fully understood and the exposure needs to be better characterized to improve the basis for risk assessment in occupational and environmental settings. Part of this knowledge gap was filled by a series of controlled human diesel exhaust exposure studies carried out in Umeå, Sweden. They monitored exposure, and

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investigated the mechanisms behind the observed effects. Healthy and asthmatic volunteers were exposed in an exposure chamber to either DE concentrations of 300 $\mu\text{g}/\text{m}^3$ (measured as PM_{10}) for 1 h [17-19] or 100 $\mu\text{g}/\text{m}^3$ for 2 h [20,21]. The results demonstrated that DE exposure caused inflammatory responses at 6 h and 18 h. This was indicated by increased cell counts (neutrophils, B lymphocytes and eosinophils) and concentrations of fibronectin and methyl histamine in airway lining fluid, as well as increase neutrophils and mast cells, up-regulation of endothelial adhesion molecules and interleukins in bronchial biopsies [17-19,22]. In one experiment, this airway inflammation was also followed by an increase of neutrophils and platelets in peripheral blood [19].

Additionally, a DE-induced bronchoconstrictive response (increased airway resistance and specific airway resistance) was observed as measured by computerized whole body plethysmography [20,23,24]. However, no significant change was detected with conventional pulmonary function tests (dynamic spirometry and peak expiratory flow) [19,25].

The Umeå studies have provided extensive knowledge on the respiratory effects of moderate DE exposure. However, the absence of lung function changes in conventional tests in most of the Umeå studies may be due to the relatively short exposure time and follow-up time. In the present investigation, which is a part of the DINO study "Health Effects of Combined Exposure to Diesel and Noise", we prolonged the exposure time to 3 h and monitored symptoms and peak expiratory flow (PEF) over the whole exposure period. The aim was to find potential relationships between exposure and effects, and to monitor how quickly such changes would develop.

Materials and methods

Subjects

Eighteen healthy, non-smoking volunteers (9 males and 9 females) with a mean age of 51 (range 40–66) years were recruited. Inclusion criteria were no symptoms or diagnosis of asthma, normal chest auscultation, lung function and a negative skin prick test to a standard panel of aeroallergens for atopy. The study was approved by the Regional Ethical Review Board, and all subjects provided written consent.

Exposure protocol

Each subject underwent four different exposure scenarios at rest (seated quietly): (1) Reference exposure: low DE concentration (measured as mass concentration of particles with an aerodynamic diameter less than 1 μm , PM_{10} ~ 2 $\mu\text{g}/\text{m}^3$) and low traffic noise (46 dB(A)); (2) Diesel exposure: high DE concentration (PM_{10} ~ 300 $\mu\text{g}/\text{m}^3$) and low traffic noise; (3) Noise exposure: low DE concentration and high traffic noise (75 dB(A)); and (4) Diesel and noise exposure: high DE concentration and high traffic noise. Exposure lasted for 3 h and took place in a specially built exposure chamber

(22 m^2 , suitable for 3 persons simultaneously) on different occasions in a randomized sequence, at least one week apart. The study design was double-blind, however, the difference between high and low traffic noise was easily noticed by study subjects and the physician.

In this paper, the reference exposure and the noise exposure were merged and renamed "filtered air" (FA) exposure, while the diesel exposure and the diesel and noise exposure were merged and renamed "diesel exhaust" (DE) exposure, since no interactions were found between DE and noise in any marker of respiratory effects (see Statistical analysis section).

The DE was generated from a passenger car (Volkswagen Passat TDI, -98, 1900 cm^3 , 81 kW) running in idle mode. Swedish Environmental Class 1 diesel fuel with a sulfur content less than 10 ppm was used. Dilution was controlled to supply the desired concentration of particles to the chamber for the different exposure scenarios. Details of diesel aerosol generation, as well as DE characteristics in both particle and gas phase are reported elsewhere [26]. Traffic noise recorded at a street crossing was played from two loudspeakers reproducing each channel in a stereo recording. The noise had a continuous, natural change of noise level. The temperature and the relative humidity in the chamber were maintained at 23°C and between 30% and 40%, respectively. Characteristics of DE and FA exposures are presented in Table 1.

Self-reported symptoms and medical examinations

The questions about symptoms of upper airways, including runny nose, nasal congestion, throat irritation and chest tightness, were rated on a visual analogue scale (VAS) (range 0 to 100 millimeter) before exposure, and at 15, 75 and 135 min into exposure during each exposure session. The VAS is most appropriate at repeated measurement within subjects as it has the sensitivity required to measure minute changes [27]. Signs of nose and throat irritation (redness/secretion/swelling) were recorded (normal/slight/moderate/pronounced), and auscultation of lungs was performed in a medical examination before and after exposure by the same

Table 1 Exposure details during DE and FA exposures

	DE exposure	FA exposure
PM_{10} mass concentration ($\mu\text{g}/\text{m}^3$)	276 \pm 27	2 \pm 2
Number concentration (particles/ cm^3)	388000 \pm 48000	14 \pm 16
NO (ppm)	9.8 \pm 1.0	0.003 \pm 0.001
NO_2 (ppm)	1.3 \pm 0.4	0.002 \pm 0.001
CO_2 (ppm)	1945 \pm 125	947 \pm 67
Formaldehyde ($\mu\text{g}/\text{m}^3$)	400 \pm 97	7.5
Temperature (°C)	22.7 \pm 0.7	22.7 \pm 0.8

Values are mean \pm standard deviation (SD).
DE, diesel exhaust; FA, filtered air.

physician. For scheduling of examinations and samplings see Table 2.

Lung function and nasal patency

PEF was measured in two ways: (1) PEF data output from a Mini-Wright Peak Flow Meter (MWPFM) with a measuring range of 60–800 L/min, which was measured four times (three recordings each time) in each exposure session: before exposure and at 15, 75 and 135 min into exposure; (2) PEF data output from a computerized spirometer (SPIRARE 3, DIAGNOSTICA, Oslo, Norway), which was measured only before and after exposure (three recordings each time).

Rhinometry and spirometry (a minimum of three recordings for each outcome) were performed before and after exposure: forced vital capacity (FVC) and forced expiratory volume in one second (FEV₁) were measured according to the European Respiratory Society guidelines [28], using the computerized spirometer. The minimal cross-sectional area between 0–22 mm (MCA₁) and between 22–54 mm (MCA₂) into the nasal opening, and the volume of the nasal cavity between these distances (Volume₁ and Volume₂) were determined using acoustic rhinometry (RhinoScan v. 2.5, Interacoustics A/S, Assens, Denmark) [29]. The “best” results at each time point from PEF, spirometry and acoustic rhinometry measurements were included in the statistical analysis.

Nasal lavage

Nasal lavage was performed in this study since it has been proposed as a noninvasive method to detect upper airway inflammation of air pollutants [30,31]. We conducted nasal lavage before exposure, after exposure, and at 20 h post-exposure (range 18–22 h), with minor modifications as described by Benson et al. [32]. Briefly, 18 ml 37°C 0.9% NaCl solution was instilled with a syringe into one nostril until the liquid appeared in the opposite one. The liquid was sucked back and instilled again. The procedure was repeated three times in each nostril. The lavage liquid was immediately chilled and the cells were separated by centrifugation (Centrifuge 5702R, Eppendorf

AG, Hamburg, Germany). The supernatant was separated from the pellet and stored at –80°C until analysis of inflammatory markers.

Inflammatory markers in blood and nasal lavage

Peripheral blood samples (before exposure, after exposure, 20 h post-exposure) were analyzed for total and differential leukocyte cell counts (Sysmex XE-1800i). Plasma fibrinogen (Sysmex CA-7000) and C reactive protein (CRP, Cobas c501) concentrations were determined.

The serum concentrations of interleukin-6 (IL-6) and IL-8, and the nasal lavage concentrations of IL-6, IL-8, intercellular adhesion molecule-1 (ICAM-1) and tumor necrosis factor- α (TNF- α) were analyzed by multiplexed magnetic bead-based immunoassays on a Luminex platform (Bio-Plex 200, Bio-Rad Life Science, Hercules, CA).

Clara cell protein (CC16) and surfactant protein D (SP-D) in serum were analyzed using commercial ELISA kits from Biovendor (BioVendor Laboratory Medicine, Inc., Brno, Czech Republic), according to the protocols supplied by the manufacturer. The imprecision, expressed as coefficient of variation, was 13% for CC16 and 12% for SP-D, as calculated from duplicate analyses.

Statistical analysis

Absolute changes from baseline were calculated for each individual and each exposure scenario to reduce the influence of inter-individual variation for the data from self-rated symptoms, lung function, biomarkers in blood and nasal lavage. Then the four exposure scenarios were merged into two, as explained above, since the effect of noise did not modify the effect of DE in any of the markers ($p > 0.06$ for all); the variation of the mean PM₁ mass concentrations between all DE exposures (i.e. the diesel exposures and the diesel and noise exposures) from the mean value of 276 $\mu\text{g}/\text{m}^3$ (Table 1) was on average smaller than 6.5%. The changes in the selected outcome measures at DE exposure vs. changes at FA exposure were analyzed with repeated-measures analysis of variance using a linear model type of the generalized estimating equation in SPSS 18.0 (SPSS Inc., Chicago,

Table 2 Scheduling and time point of different examinations, blood and nasal lavage samplings

Item	Before exposure (outside chamber)	During exposure (in chamber)			After exposure (outside chamber)	20 h post-exposure (outside chamber)
		15 min	75 min	135 min		
Self-rating symptoms	× (7:40)	× (9:45)	× (10:45)	× (11:45)		
Medical examination	× (7:00)				× (13:10)	
Peak expiratory flow	× (7:40)	× (9:45)	× (10:45)	× (11:45)		
Rhinometry	× (8:30~9:00)				× (13:20~13:50)	
Spirometry	× (8:30~9:00)				× (13:20~13:50)	
Blood samples	× (8:30~9:00)				× (13:20~13:50)	× (7:00~8:30)
Nasal lavage	× (8:30~9:00)				× (13:20~13:50)	× (7:00~8:30)

IL, USA). Subject identification, the four initial exposure scenarios, and the time point of measurements were used to indicate the repeated measurements. Exposure sequence was not included in the final models since inclusion of the exposure sequence did not change the results.

The data from the medical examination was transformed to binary variables. If signs (redness/secretion/swelling of nose, throat, and sound of lung and heart auscultation) after exposure became worse than before exposure (from normal to slight, and so on), we recorded it as “getting worse”, otherwise “no change”. Any “getting worse” sign of nose and throat, or sound of lungs represented a corresponding positive finding of upper-airway irritation or lung signs. A binary logistic model type of generalized estimating equation was used to estimate the odds ratios.

Statistical significance refers to $p < 0.05$ (two-tailed). However, multiple comparisons were performed for self-rated symptoms (three times), PEF from MWPFM (three times), and inflammatory markers in blood and nasal lavage (two times). For those comparisons, statistical significance refers to $p < 0.017$ and $p < 0.025$, respectively, according to the Bonferroni correction.

Results

Symptoms and Signs

Scores of self-rated symptoms are presented in Table 3. Subjects reported a mild but progressive irritation in the throat during DE exposure, the increment on the VAS scale was 3.75 in DE exposure versus -1.53 in FA exposure ($p = 0.016$) at 135 min. No significant differences between exposures were detected in self-rated nose or chest symptoms.

Irritation in upper airways (nose and throat) was found in the physical examination. Exposure to DE increased the odds ratio (OR) of signs in the upper-airways (OR=3.2, 95% CI 1.9-11.0). No change was found in lung sounds (data not shown).

Lung function

Similar PEF values at baseline were obtained with the MWPFM and the spirometer for both FA and DE exposure (Table 4). However, after 15 min of exposure, PEF decreased during DE exposure but increased during FA exposure, and this pattern lasted throughout the whole exposure period. The difference between PEF changes in DE and FA was 14 L/min at 75 min of exposure (MWPFM, $p = 0.005$), 12 L/min at 135 min of exposure (MWPFM, $p = 0.020$) and 10 L/min after exposure (spirometer, $p = 0.078$) (Table 4).

The changes in spirometry (FEV₁, FVC) and rhinometry results were small and similar after both DE and FA exposure (Table 5).

Inflammatory markers

There was a significantly greater increment of total leukocytes in blood at 20 h post-exposure following DE exposure compared to FA exposure (Table 6, $p = 0.007$), including a significantly higher increment of monocyte counts ($p = 0.017$). A trend towards an increase in monocyte counts between DE exposure and FA exposure was suggested also immediately after exposure ($p = 0.04$). No such patterns were discerned for neutrophils, eosinophils or lymphocytes.

Data on other inflammatory markers in serum and nasal lavage are shown in Table 7. We only found a significant difference between CC16 changes in DE and FA exposure. A slight increase in CC16 was observed immediately after DE exposure and returned to baseline at 20 h post-exposure for DE exposure, while it increased at 20 h post-exposure of FA exposure ($p = 0.016$). A trend towards a greater increment of serum IL-6 for DE exposure than FA exposure was suggested at 20 h post-exposure ($p = 0.066$). However, CRP tended to decrease more rapidly after exposure to DE than after FA ($p = 0.034$).

Table 3 Effects of FA exposure and DE exposure on self-rated symptoms

	Before exposure	15 min exposure	75 min exposure	135 min exposure	Δ 15 min ^a	Δ 75 min ^a	Δ 135 min ^a
FA exposure							
Runny nose	1.39±7.83	0.08±0.5	0.14±0.83	0.00±0.00	-1.31±7.86	-1.25±7.88	-1.39±7.83
Nasal congestion	1.69±7.99	0.53±1.18	0.61±1.87	0.36±1.36	-1.17±8.12	-1.08±8.21	-1.33±8.11
Throat irritation	1.66±7.46	1.00±5.01	0.11±0.46	0.08±0.50	-0.61±8.64	-1.50±7.03	-1.53±7.36
Chest tightness	-- [*]	-- [*]	-- [*]	0.14±0.83	-- [*]	-- [*]	0.14±0.83
DE exposure							
Runny nose	2.86±10.91	1.97±10.09	1.44±7.66	1.69±8.34	-0.89±3.44	-1.42±4.86	-1.17±4.31
Nasal congestion	3.03±10.55	1.72±7.52	2.69±8.08	3.53±9.32	-1.31±4.33	-0.33±8.10	0.50±6.98
Throat irritation	2.31±6.96	1.06±2.27	4.00±8.64	6.06±12.11	-1.25±6.21	1.69±10.32	3.75±12.11 [#]
Chest tightness	0.28±1.67	0.17±0.74	0.17±0.74	0.56±2.29	-0.11±1.51	-0.11±1.85	0.28±2.88

Values are mean ± standard deviation (SD) using a visual analog scale.

DE, diesel exhaust; FA, filtered air.

^aΔ 15 min = 15 min of exposure - before exposure; Δ 75 min = 75 min of exposure - before exposure; Δ 135 min = 135 min of exposure - before exposure.

[#]No symptoms were reported. ^{*} $P < 0.017$ for DE exposure vs. FA exposure.

Table 4 Effects of FA exposure and DE exposure on PEF as measured using a MWPFM and a SPIRARE spirometer

	Before exposure	15 min exposure	75 min exposure	135 min exposure	After exposure	Δ 15 min ^a	Δ 75 min ^a	Δ 135 min ^a	Δ after exposure ^a
FA exposure									
PEF (L/min) from MWPFM	530±73	533±73	534±73	538±75	--*	3±25	4±26	8±31	--*
PEF (L/min) from spirometer	524±88	--*	--*	--*	531±90	--*	--*	--*	8±27
DE exposure									
PEF (L/min) from MWPFM	537±74	531±75	528±78	534±81	--*	-6±22	-10±22 [#]	-4±18	--*
PEF (L/min) from spirometer	530±93	--*	--*	--*	527±89	--*	--*	--*	-3±24

Values are mean ± standard deviation (SD).

DE, diesel exhaust; FA, filtered air; PEF, peak expiratory flow; MWPFM, Mini-Wright Peak Flow Meter.

^aΔ 15 min = 15 min of exposure – before exposure; Δ 75 min = 75 min of exposure – before exposure; Δ 135 min = 135 min of exposure – before exposure;

Δ after exposure = after exposure – before exposure.

[#]No PEF was recorded. *P <0.017 for DE exposure vs. FA exposure.

Discussion

This study was extracted from DINO project, the overall aim of which was to evaluate the combined effect of DE and traffic noise on human physiological response (including also cardiovascular responses such as heart rate variability). Although effects of traffic noise on airways have not been documented, we still initially included traffic noise in the analysis, to see if it modified the reported or observed airway effects of DE. However, no such modifying effect was found in any markers of respiratory response. Moreover, the generation of DE was highly reproducible between the sessions. Thus, for optimal statistical power, we merged the four exposures into two: exposure or no exposure to DE.

The concentration of DE in this study was nearly 300 µg/m³, and such levels may occur at rush hours in large city centers. We found divergent trends in PEF changes during DE and FA exposure, and a significant DE-associated PEF decrease at 75 min in healthy subjects. Self-reported symptoms and the physical examination indicated irritation of the upper airways. The increase in peripheral leukocyte counts suggests a mild systemic inflammatory response, supported also by a tendency towards an increase in serum IL-6.

To the best of our knowledge, this is the first study that shows an effect on PEF in healthy subjects. DE induced a slight but consistent reduction in PEF that was significantly different from FA exposure at 75 min. Thereafter

Table 5 Effects of FA exposure and DE exposure on spirometry (FEV₁, FVC) and rhinometry (MCA₁, MCA₂, Volume₁, Volume₂)

		Before exposure	After exposure	Δ after exposure ^a
Spirometry				
FEV ₁	FA exposure	3.23±0.57	3.26±0.58	0.02±0.13
	DE exposure	3.23±0.60	3.26±0.59	0.03±0.08
FVC	FA exposure	4.12±0.71	4.14±0.70	0.02±0.16
	DE exposure	4.13±0.72	4.17±0.72	0.04±0.10
Rhinometry				
MCA ₁	FA exposure	1.12±0.28	1.13±0.23	0.01±0.18
	DE exposure	1.08±0.24	1.13±0.24	0.05±0.16
MCA ₂	FA exposure	0.95±0.23	1.06±0.25	0.11±0.19
	DE exposure	0.94±0.20	1.03±0.31	0.10±0.23
Volume ₁	FA exposure	4.06±0.75	4.07±0.67	0.01±0.26
	DE exposure	4.05±0.72	4.10±0.69	0.05±0.21
Volume ₂	FA exposure	6.90±1.56	8.16±1.59	1.26±1.74
	DE exposure	6.51±1.19	7.82±1.93	1.32±1.58

Values are mean ± standard deviation (SD).

DE, diesel exhaust; FA, filtered air; FEV₁, forced expiratory volume in one second; FVC, forced vital capacity; MCA₁, minimal cross-sectional area between 0–22 mm; MCA₂, minimal cross-sectional area between 22–45 mm.

^aΔ after exposure = after exposure – before exposure.

Table 6 Effects of FA exposure and DE exposure on cell count in peripheral blood

Cell type	Cell count ($\times 10^9/L$)					P-values for DE exposure vs. FA exposure	
	Before exposure	After exposure	20 h post-exposure	Δ after exposure ^a	Δ 20 h post-exposure ^a	Δ after exposure ^a	Δ 20 h post-exposure ^a
FA exposure							
Leukocytes	5.45 \pm 1.36	5.72 \pm 1.34	5.17 \pm 1.17	0.27 \pm 0.53	-0.28 \pm 1.02	-	-
Neutrophils	3.02 \pm 1.15	2.99 \pm 1.03	2.64 \pm 0.75	-0.04 \pm 0.31	-0.38 \pm 0.99	-	-
Eosinophils	0.19 \pm 0.09	0.20 \pm 0.11	0.21 \pm 0.11	0.01 \pm 0.06	0.02 \pm 0.06	-	-
Lymphocytes	1.69 \pm 0.49	2.02 \pm 0.64	1.83 \pm 0.53	0.33 \pm 0.32	0.14 \pm 0.30	-	-
Monocytes	0.49 \pm 0.13	0.48 \pm 0.15	0.44 \pm 0.12	-0.01 \pm 0.13	-0.05 \pm 0.11	-	-
DE exposure							
Leukocytes	5.29 \pm 1.00	5.60 \pm 0.98	5.45 \pm 1.37	0.31 \pm 0.55	0.16 \pm 1.01	0.725	0.007
Neutrophils	3.00 \pm 0.81	2.91 \pm 0.63	2.89 \pm 1.04	-0.09 \pm 0.44	-0.11 \pm 0.93	0.474	0.139
Eosinophils	0.19 \pm 0.12	0.20 \pm 0.13	0.22 \pm 0.13	0.01 \pm 0.06	0.03 \pm 0.07	0.847	0.431
Lymphocytes	1.59 \pm 0.48	1.94 \pm 0.59	1.82 \pm 0.64	0.35 \pm 0.24	0.23 \pm 0.35	0.651	0.264
Monocytes	0.47 \pm 0.11	0.51 \pm 0.10	0.48 \pm 0.13	0.04 \pm 0.11	0.01 \pm 0.10	0.042	0.017

Values are mean \pm standard deviation (SD).

DE, diesel exhaust; FA, filtered air.

^a Δ after exposure = after exposure - before exposure; Δ 20 h post-exposure = 20 h post-exposure - before exposure.

differences were still suggested at 135 min and after exposure, approaching significance. Our interpretation is that DE caused a transient lung function change with rapid recovery. PEF was seldom measured at DE exposure in healthy volunteers, but in asthmatics exposed to DE (PM₁₀ 100 $\mu\text{g}/\text{m}^3$) for 2 h, there was a non-significant trend towards a decreased PEF immediately after exposure [33]. Similar to other studies [20,34], we performed spirometry before and after exposures and failed to find significant difference in FEV₁ changes, although we used a longer exposure time (3 h). This also reflected that the time points of monitoring may be important. In this study, there was a 50 min delay between end of exposure and spirometry, which may allow the lung function to recover. Moreover, the subjects of this study were older (mean age about 50 years) than in other studies.

The small but significant PEF changes in healthy subjects may imply that occupational DE limit values should be reconsidered. DE is quite common in occupational settings, and some studies have indicated adverse effects on lung function at long-term occupational exposure to DE in combination with dust [10,35]. The current American Conference of Governmental Industrial Hygienists (ACGIH) Threshold Limit Values (TLVs) for respirable particulates and NO₂ are 3 mg/m³ and 3 ppm as 8-hour time-weighted averages (TWAs), respectively [36]. However, the present study indicates that substantially lower exposures (PM₁ level of 300 $\mu\text{g}/\text{m}^3$ and NO₂ of 1.3 ppm) to DE for only three hours will cause a transient effect on lung function.

As in previous studies [23,37], self-reported throat and eye irritation [26] increased during DE exposure, in agreement with objective signs at the physical examination. The emission of aldehydes is the most likely cause in the present study. A Swedish study found an increase in nasal irritation and lower respiratory tract symptoms at an average exposure of 260 $\mu\text{g}/\text{m}^3$ of formaldehyde [38]. Our exposure level of formaldehyde, about 400 $\mu\text{g}/\text{m}^3$, was higher than in these reports, and DE also contains other aldehydes such as acrolein that may cause such irritant symptoms.

In the present study, we used nasal lavage for measurement of the acute cytokines (IL-6, IL-8, TNF- α , ICAM-1) to define the timing of inflammatory response in the nasal cavity instead of the more invasive bronchoalveolar lavage and bronchoscopy techniques. However, such repeated sampling of airway fluid could have the drawback of washout effect [30,39], resulting in lower concentration of biomarkers in subsequent lavages and then diminishing real differences between DE and FA exposures. The only finding in nasal lavage in this study was a slight increase (not statistically significant) of IL-6 after DE exposure at 20 h post-exposure. Other cytokines (IL-8, TNF- α , ICAM-1), on the contrary, remained unchanged. Nasal lavage has not been widely investigated in chamber studies of DE exposure, Blomberg et al. [40] only found an increase in ascorbic acid concentration in nasal lavage after diesel exposure (1 h, PM 300 $\mu\text{g}/\text{m}^3$). In other in vivo studies, intranasal challenge with allergen + DE in atopic subjects was found to increase IgE, IL-4 and histamine

Table 7 Effects of FA exposure and DE exposure on inflammatory markers

Inflammatory markers	Concentration of markers					P-values for DE exposure vs. FA exposure	
	Before exposure	After exposure	20 h post-exposure	Δ after exposure ^a	Δ 20 h post-exposure ^a	Δ after exposure ^a	Δ 20 h post-exposure ^a
FA exposure							
Serum IL-6 (pg/ml)	1.73±0.98	1.69±1.13	1.60±1.33	-0.04±0.56	-0.13±0.90	-	-
Serum IL-8 (pg/ml)	5.00±3.28	4.91±3.26	5.46±4.35	-0.09±1.33	0.46±2.48	-	-
NL IL-6 (pg/ml)	0.97±1.01	0.94±0.86	0.85±0.68	-0.03±0.93	-0.11±0.81	-	-
NL IL-8 (pg/ml)	67.0±73.3	48.5±50.5	49.9±66.6	-18.6±71.5	-17.1±49.4	-	-
NL TNF- α (pg/ml)	0.95±0.77	0.84±0.14	1.03±0.88	-0.10±0.79	0.08±1.19	-	-
NL ICAM-1(pg/ml)	76.8±87.1	76.6±101	62.0±73.9	-0.15±114	-14.7±64.0	-	-
CRP (mg/l)	1.6±1.6	1.5±1.7	1.3±1.2	-0.03±0.23	-0.27±0.54	-	-
Fibrinogen (g/l)	2.9±0.5	2.9±0.5	3.1±0.6	0.12±0.72	0.24±0.66	-	-
CC16 (μ g/l)	5.93±1.98	5.84±2.20	6.73±2.38	-0.08±0.99	0.61±1.56	-	-
SP-D (μ g/l)	66.9±34.0	66.7±35.0	70.8±36.1	-0.21±9.42	3.90±9.81	-	-
DE exposure							
Serum IL-6 (pg/ml)	1.80±1.35	1.64±1.14	1.92±1.56	-0.16±0.64	0.12±0.78	0.413	0.066
Serum IL-8 (pg/ml)	5.10±3.62	4.85±3.79	5.59±3.74	-0.25±1.90	0.49±1.82	0.666	0.940
NL IL-6 (pg/ml)	1.32±2.03	1.29±2.08	2.01±5.04	-0.03±2.35	0.70±4.64	0.983	0.247
NL IL-8 (pg/ml)	71.3±71.7	63.9±97.8	48.6±46.7	-7.40±98.4	-22.7±48.1	0.483	0.597
NL TNF- α (pg/ml)	0.82±0.00	0.90±0.37	0.98±0.79	0.08±0.37	0.16±0.79	0.312	0.715
NL ICAM-1(pg/ml)	109.7±138	93.5±142	85.3±103	-16.2±114	-24.4±74.5	0.409	0.398
CRP (mg/L)	2.0±2.2	1.9±2.0	1.6±1.6	-0.12±0.24	-0.39±0.71	0.034	0.283
Fibrinogen (g/l)	3.2±0.7	3.3±0.8	3.4±0.10	0.13±0.55	0.26±0.84	0.968	0.940
CC16 (μ g/l)	6.21±2.23	6.64±3.44	6.12±2.32	0.43±2.50	-0.09±1.44	0.224	0.016
SP-D (μ g/l)	63.4±34.0	61.9±32.7	66.7±32.8	-1.45±4.78	3.34±12.1	0.626	0.838

Values are mean \pm standard deviation (SD).

DE, diesel exhaust; FA, filtered air; NL, nasal lavage.

^a Δ after exposure = after exposure - before exposure; Δ 20 h post-exposure = 20 h post-exposure - before exposure.

concentrations in nasal lavage [41,42], but concentrations of IL-6, IL-8 were not altered by intranasal challenge with DE alone in healthy volunteers [43].

CC16 is an anti-inflammatory protein secreted by the Clara cells located in the bronchioli [44], and the transfer to serum is increased when the permeability of the lung epithelial barrier is compromised [45]. Two chamber studies found increased serum CC16 after exposure to wood smoke at PM mass concentrations similar to those in the present study [46,47]. CC16 levels were, however, seldom reported at DE exposure in previous chamber studies. In this study, the slight increase of serum CC16 immediately after DE exposure was not statistically significant and levels returned to baseline at 20 h post-exposure. The reason for the increase in serum CC16 the morning after FA exposure is unclear.

DE exposure, at the moderate levels used in this study, induced a systemic inflammatory response in peripheral

blood. Previous experimental studies showed white cell and proinflammatory cytokines changes in bronchoalveolar lavage and bronchial biopsies, but rarely in peripheral blood. Total leukocyte counts were higher after 3 h exposure to both DE and FA in the afternoon than before exposure in the morning, which reflects the normal circadian variation [48]. However, the total leukocyte counts remained increased at 20 h post-exposure after DE exposure, but not after FA exposure, which suggests a slight systemic cellular inflammatory response after exposed to DE. Changes in pro-inflammatory cytokines in peripheral blood were also identified. The trend towards an increase of serum IL-6 ($p=0.07$) after DE exposure at 20 h post-exposure, which was also observed by Tornqvist et al. [49], may represent a pro-inflammatory systemic response. However, consistent with other controlled DE chamber studies performed in either healthy volunteers [50,51] or individuals with chronic obstructive pulmonary

disease [52], we did not find a DE-associated increase in CRP. The follow-up time in the present study perhaps has been too short to detect a change in CRP.

Conclusion

In summary, this study shows that short-term exposure to DE at 300 $\mu\text{g}/\text{m}^3$ can cause irritation in upper airways, along with a temporary decline in PEF in healthy subjects, after only 75 min into exposure. Additionally, the increase in leukocyte cell counts in peripheral blood indicates that these levels of DE exposure also cause a systemic inflammatory response.

Abbreviations

DE: Diesel exhaust; PM: Particulate matter; PEF: Peak expiratory flow; FA: Filtered air; VAS: Visual analogue scale; MWPFM: Mini-Wright Peak Flow Meter; FVC: Forced vital capacity; FEV₁: Forced expiratory volume in one second; MCA₁: Minimal cross-sectional area between 0–22 mm; MCA₂: Minimal cross-sectional area between 22–45 mm; CRP: C reactive protein; IL: Interleukin; ICAM: Intercellular adhesion molecule; TNF: Tumor necrosis factor; CC16: Clara cell protein; SP-D: Surfactant protein D; OR: Odds ratio; ACGIH: American Conference of Governmental Industrial Hygienists; TLVs: Threshold Limit Values; TWAs: 8-hour time-weighted averages.

Competing interests

The authors declare that they have no competing financial interests

Authors' contributions

AW, AG and MA designed the project and performed the chamber exposure. AG and AW generated the aerosol. AW provided exposure characteristics. JN and MA were responsible for the medical examinations. BJ and MK measured biomarkers. YX, LB, AA, MK and MA analyzed the data and interpreted the results. The manuscript was written by YX and revised critically by LB and MA. All authors read, corrected and approved the manuscript.

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References

1. de-Kok TM, Drieste HA, Hogervorst JG, Briede JJ: **Toxicological assessment of ambient and traffic-related particulate matter: a review of recent studies.** *Mutat Res* 2006, **613**:103–122.

2. Pope CA 3rd, Dockery DW: **Health effects of fine particulate air pollution: lines that connect.** *J Air Waste Manag Assoc* 2006, **56**:709–742.
3. Brook RD, Rajagopalan S, Pope CA 3rd, Brook JR, Bhatnagar A, Diez-Roux AV, Holguin F, Hong Y, Luepker RV, Mittleman MA, Peters A, Siscovick D, Smith SC Jr, Whitsett L, Kaufman JD: **Particulate matter air pollution and cardiovascular disease: an update to the scientific statement from the American Heart Association.** *Circulation* 2010, **121**:2331–2378.
4. Gan WQ, Fitzgerald JM, Carlsten C, Sadatsafavi M, Brauer M: **Associations of ambient air pollution with chronic obstructive pulmonary disease hospitalization and mortality.** *Am J Respir Crit Care Med* 2013, **187**(7):721–7.
5. Faustini A, Stafoggia M, Colais P, Berti G, Bisanti L, Cadum E, Cernigliaro A, Mallone S, Scarnato C, Forastiere F: **Air pollution and multiple acute respiratory outcomes.** *Eur Respir J* 2013. Epub ahead of print.
6. World Health Organization: **Burden of disease associated with urban outdoor air pollution for 2008; 2008.** http://www.who.int/phe/health_topics/outdoorair/databases/burden_disease/en/index.html.
7. U.S. Environmental Protection Agency: **Health assessment document for diesel engine exhaust.** Report EPA/600/8-90/057F. Washington, DC: U.S. Environmental Protection Agency; 2002.
8. U.S. Environmental Protection Agency: **Air quality criteria for particulate matter (October 2004) Volume I.** Report EPA/600/p-99/002AF. Washington, DC: U.S. Environmental Protection Agency; 2004.
9. Gauderman WJ, Vora H, McConnell R, Berhane K, Gilliland F, Thomas D, Lurmann F, Avol E, Kunzli N, Jerrett M, Peters J: **Effect of exposure to traffic on lung development from 10 to 18 years of age: a cohort study.** *Lancet* 2007, **369**:571–577.
10. Ulvestad B, Bakke B, Eduard W, Kongerud J, Lund MB: **Cumulative exposure to dust causes accelerated decline in lung function in tunnel workers.** *Occup Environ Med* 2001, **58**:663–669.
11. Riedl M, Diaz-Sanchez D: **Biology of diesel exhaust effects on respiratory function.** *J Allergy Clin Immunol* 2005, **115**:221–228. quiz 229.
12. Hart JE, Laden F, Eisen EA, Smith TJ, Garshick E: **Chronic obstructive pulmonary disease mortality in railroad workers.** *Occup Environ Med* 2009, **66**:221–226.
13. Hart JE, Eisen EA, Laden F: **Occupational diesel exhaust exposure as a risk factor for chronic obstructive pulmonary disease.** *Curr Opin Pulm Med* 2012, **18**:151–154.
14. Pedeli X, Hoek G, Katsouyanni K: **Risk assessment of diesel exhaust and lung cancer: combining human and animal studies after adjustment for biases in epidemiological studies.** *Environ Health* 2011, **10**:30.
15. Olsson AC, Gustavsson P, Kromhout H, Peters S, Vermeulen R, Bruske I, Pesch B, Siemiatycki J, Pintos J, Bruning T, Cassidy A, Wichmann HE, Consonni D, Landi MT, Caporaso N, Plato N, Merletti F, Mirabelli D, Richiardi L, Jockel KH, Ahrens W, Pohlabeln H, Lissowska J, Szeszenia-Dabrowska N, Zaridze D, Stucker I, Benhamou S, Bencko V, Foretova L, Janout V, Rudnai P, Fabianova E, Dumitru RS, Gross IM, Kendzia B, Forastiere F, Bueno-de-Mesquita B, Brennan P, Boffetta P, Straif K: **Exposure to diesel motor exhaust and lung cancer risk in a pooled analysis from case-control studies in Europe and Canada.** *Am J Respir Crit Care Med* 2011, **183**:941–948.
16. International Agency for Research on Cancer: **Carcinogenicity of diesel-engine and gasoline-engine exhausts and some nitroarenes.** Lyon: IARC Monographs on the Evaluation of Carcinogenic Risks to Humans Vol 125; 2012.
17. Sehlstedt M, Behndig AF, Boman C, Blomberg A, Sandstrom T, Pourazar J: **Airway inflammatory response to diesel exhaust generated at urban cycle running conditions.** *Inhal Toxicol* 2010, **22**:1144–1150.
18. Pourazar J, Frew AJ, Blomberg A, Helleday R, Kelly FJ, Wilson S, Sandstrom T: **Diesel exhaust exposure enhances the expression of il-13 in the bronchial epithelium of healthy subjects.** *Respir Med* 2004, **98**:821–825.
19. Salvi S, Blomberg A, Rudell B, Kelly F, Sandstrom T, Holgate ST, Frew AJ: **Acute inflammatory responses in the airways and peripheral blood after short-term exposure to diesel exhaust in healthy human volunteers.** *Am J Respir Crit Care Med* 1999, **159**:702–709.
20. Stenfors N, Nordenhall C, Salvi SS, Mudway I, Soderberg M, Blomberg A, Helleday R, Levin JO, Holgate ST, Kelly FJ, Frew AJ, Sandstrom T: **Different airway inflammatory responses in asthmatic and healthy humans exposed to diesel.** *Eur Respir J* 2004, **23**:82–86.
21. Behndig AF, Mudway IS, Brown JL, Stenfors N, Helleday R, Duggan ST, Wilson SJ, Boman C, Cassee FR, Frew AJ, Kelly FJ, Sandstrom T, Blomberg A: **Airway antioxidant and inflammatory responses to diesel exhaust exposure in healthy humans.** *Eur Respir J* 2006, **27**:359–365.
22. Salvi SS, Nordenhall C, Blomberg A, Rudell B, Pourazar J, Kelly FJ, Wilson S, Sandstrom T, Holgate ST, Frew AJ: **Acute exposure to diesel exhaust**

- increases il-8 and gro-alpha production in healthy human airways. *Am J Respir Crit Care Med* 2000, **161**:550–557.
23. Rudell B, Ledin MC, Hammarstrom U, Stjernberg N, Lundback B, Sandstrom T: **Effects on symptoms and lung function in humans experimentally exposed to diesel exhaust.** *Occup Environ Med* 1996, **53**:658–662.
24. Nordenhall C, Pourazar J, Ledin MC, Levin JO, Sandstrom T, Adelroth E: **Diesel exhaust enhances airway responsiveness in asthmatic subjects.** *Eur Respir J* 2001, **17**:909–915.
25. Nightingale JA, Maggs R, Cullinan P, Donnelly LE, Rogers DF, Kinnersley R, Chung KF, Barnes PJ, Ashmore M, Newman-Taylor A: **Airway inflammation after controlled exposure to diesel exhaust particulates.** *Am J Respir Crit Care Med* 2000, **162**:161–166.
26. Wierzbicka A, Nilsson P, Rissler J, Sällsten G, Xu Y, Pagels J, Albin M, Osterberg K, Strandberg B, Eriksson A, Bohgard M, Bergemalm-Rynell K, Gudmundsson A: **Detailed diesel exhaust characteristics including particle surface area and lung deposited dose for better understanding of health effects in human laboratory exposure studies.** *Atmos Environ* 2013, doi: 10.1016/j.atmosenv.2013.11.025.
27. Bausewein C, Farquhar M, Booth S, Gysels M, Higginson IJ: **Measurement of breathlessness in advanced disease: a systematic review.** *Respir Med* 2007, **101**:399–410.
28. Miller MR, Hankinson J, Brusasco V, Burgos F, Casaburi R, Coates A, Crapo R, Enright P, van der Grinten CP, Gustafsson P, Jensen R, Johnson DC, MacIntyre N, McKay R, Navajas D, Pedersen OF, Pellegrino R, Viegi G, Wanger J: **Standardisation of spirometry.** *Eur Respir J* 2005, **26**:319–338.
29. Hilberg O: **Objective measurement of nasal airway dimensions using acoustic rhinometry: Methodological and clinical aspects.** *Allergy* 2002, **57**(Suppl 70):5–39.
30. Koren HS, Hatch GE, Graham DE: **Nasal lavage as a tool in assessing acute inflammation in response to inhaled pollutants.** *Toxicology* 1990, **60**:15–25.
31. Douwes J, Wouters I, Dubbeld H, van-Zwieten L, Steerenberg P, Doekes G, Heederik D: **Upper airway inflammation assessed by nasal lavage in compost workers: a relation with bio-aerosol exposure.** *Am J Ind Med* 2000, **37**:459–468.
32. Benson M, Strannegard IL, Wennergren G, Strannegard O: **Interleukin-5 and interleukin-8 in relation to eosinophils and neutrophils in nasal fluids from school children with seasonal allergic rhinitis.** *Pediatr Allergy Immunol* 1999, **10**:178–185.
33. Behndig AF, Larsson N, Brown JL, Stenfors N, Helleday R, Duggan ST, Dove RE, Wilson SJ, Sandstrom T, Kelly FJ, Mudway IS, Blomberg A: **Proinflammatory doses of diesel exhaust in healthy subjects fail to elicit equivalent or augmented airway inflammation in subjects with asthma.** *Thorax* 2011, **66**:12–19.
34. Giles LV, Carlsten C, Koehle MS: **The effect of pre-exercise diesel exhaust exposure on cycling performance and cardio-respiratory variables.** *Inhal Toxicol* 2012, **24**:783–789.
35. Lotz G, Plitzko S, Gierke E, Tittelbach U, Kersten N, Schneider WD: **Dose–response relationships between occupational exposure to potash, diesel exhaust and nitrogen oxides and lung function: cross-sectional and longitudinal study in two salt mines.** *Int Arch Occup Environ Health* 2008, **81**:1003–1019.
36. American Conference of Governmental Industrial Hygienists: *Threshold limit values for chemical substance and physical agents and biological indices for 1995/1996.* Cincinnati: American Conference of Governmental Industrial Hygienists; 1996.
37. Rudell B, Sandstrom T, Hammarstrom U, Ledin ML, Horstedt P, Stjernberg N: **Evaluation of an exposure setup for studying effects of diesel exhaust in humans.** *Int Arch Occup Environ Health* 1994, **66**:77–83.
38. Wilhelmsson B, Holmstrom M: **Possible mechanisms of formaldehyde-induced discomfort in the upper airways.** *Scand J Work Environ Health* 1992, **18**:403–407.
39. Hauser R, Garcia-Closas M, Kelsey KT, Christiani DC: **Variability of nasal lavage polymorphonuclear leukocyte counts in unexposed subjects: its potential utility for epidemiology.** *Arch Environ Health* 1994, **49**:267–272.
40. Blomberg A, Sainsbury C, Rudell B, Frew AJ, Holgate ST, Sandstrom T, Kelly FJ: **Nasal cavity lining fluid ascorbic acid concentration increases in healthy human volunteers following short term exposure to diesel exhaust.** *Free Radic Res* 1998, **28**:59–67.
41. Diaz-Sanchez D, Penichet-Garcia M, Saxon A: **Diesel exhaust particles directly induce activated mast cells to degranulate and increase histamine levels and symptom severity.** *J Allergy Clin Immunol* 2000, **106**:1140–1146.
42. Bastain TM, Gilliland FD, Li YF, Saxon A, Diaz-Sanchez D: **Intraindividual reproducibility of nasal allergic responses to diesel exhaust particles indicates a susceptible phenotype.** *Clin Immunol* 2003, **109**:130–136.
43. Kongerud J, Madden MC, Hazucha M, Peden D: **Nasal responses in asthmatic and nonasthmatic subjects following exposure to diesel exhaust particles.** *Inhal Toxicol* 2006, **18**:589–594.
44. Broecker F, Clippe A, Knoops B, Hermans C, Bernard A: **Clara cell secretory protein (cc16): features as a peripheral lung biomarker.** *Ann N Y Acad Sci* 2000, **923**:68–77.
45. Barregard L, Sällsten G, Gustafson P, Andersson L, Johansson L, Basu S, Stigendal L: **Experimental exposure to wood-smoke particles in healthy humans: effects on markers of inflammation, coagulation, and lipid peroxidation.** *Inhal Toxicol* 2006, **18**:845–853.
46. Stockfelt L, Sällsten G, Olin AC, Almerud P, Samuelsson L, Johannesson S, Molnar P, Strandberg B, Almstrand AC, Bergemalm-Rynell K: **Effects on airways of short-term exposure to two kinds of wood smoke in a chamber study of healthy humans.** *Inhal Toxicol* 2012, **24**:47–59.
47. Barregard L, Sällsten G, Andersson L, Almstrand AC, Gustafson P, Andersson M, Olin AC: **Experimental exposure to wood smoke: effects on airway inflammation and oxidative stress.** *Occup Environ Med* 2008, **65**:319.
48. Bertouch JV, Roberts-Thomson PJ, Bradley J: **Diurnal variation of lymphocyte subsets identified by monoclonal antibodies.** *Br Med J (Clin Res Ed)* 1983, **286**:1171–1172.
49. Tornqvist H, Mills NL, Gonzalez M, Miller MR, Robinson SD, Megson IL, MacNee W, Donaldson K, Soderberg S, Newby DE: **Persistent endothelial dysfunction following diesel exhaust inhalation in man.** *Am J Respir Crit Care Med* 2007, **176**:395–400.
50. Mills NL, Tornqvist H, Robinson SD, Gonzalez M, Darnley K, MacNee W, Boon NA, Donaldson K, Blomberg A, Sandstrom T, Newby DE: **Diesel exhaust inhalation causes vascular dysfunction and impaired endogenous fibrinolysis.** *Circulation* 2005, **112**:3930–3936.
51. Carlsten C, Kaufman JD, Peretz A, Trenga CA, Sheppard L, Sullivan JH: **Coagulation markers in healthy human subjects exposed to diesel exhaust.** *Thromb Res* 2007, **120**:849–855.
52. Blomberg A, Tornqvist H, Desmyter L, Deneys V, Hermans C: **Exposure to diesel exhaust nanoparticles does not induce blood hypercoagulability in an at-risk population.** *J Thromb Haemost: JTH* 2005, **3**:2103–2105.

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